

# Serial femtosecond crystallography (SFX) using X-ray free electron laser (XFEL)

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# Outline

- !! Introduction
- !! Serial femtosecond crystallography (SFX)
- !! Nanocrystal characterization and growth
- !! Nanocrystal injection and delivery methods
- !! Time-resolved studies

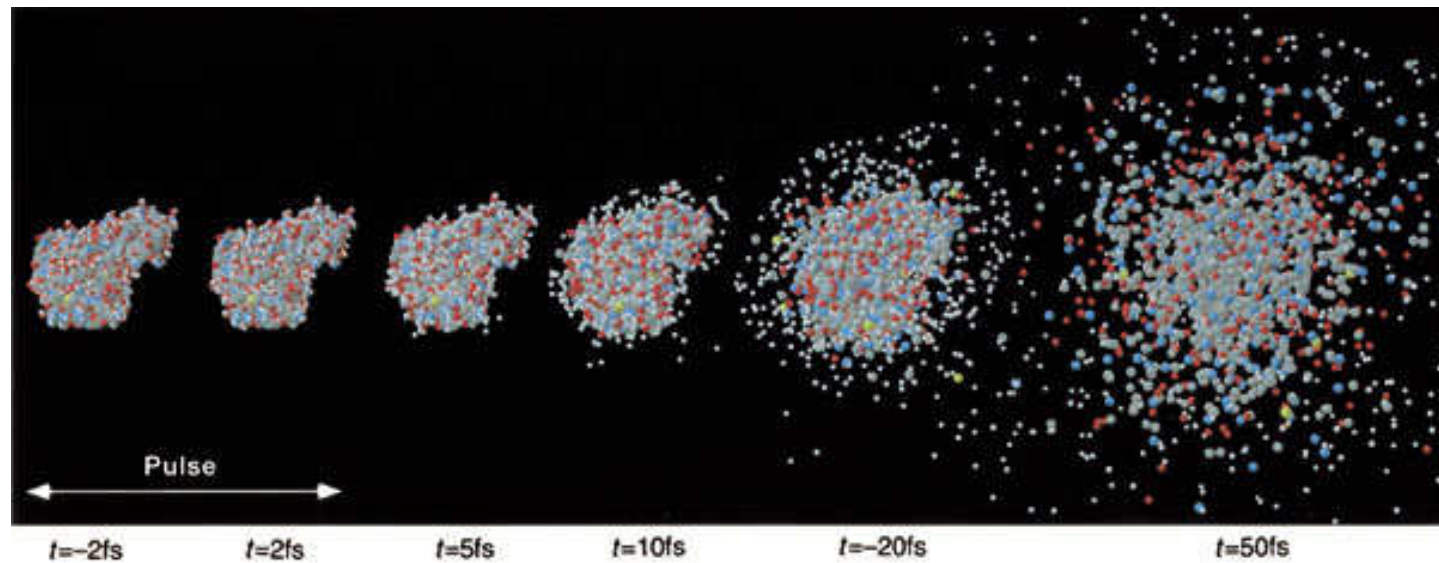
# X-ray free electron lasers (XFEL) applied to structural biology

- !! X-ray crystallography is the most applied method for the structural analysis of proteins
- !! Major limitations in protein X-ray crystallography:
  - ! Obtaining crystals of sufficient size
  - ! Radiation damage
- !! The invention of femtosecond X-ray free electron laser opens new possibilities
  - ! Nano- and Microcrystals can be used
  - ! Femtosecond X-ray pulses outrun radiation damage processes



“diffraction before destruction” principle

# Prediction of the “diffraction before destruction” principle

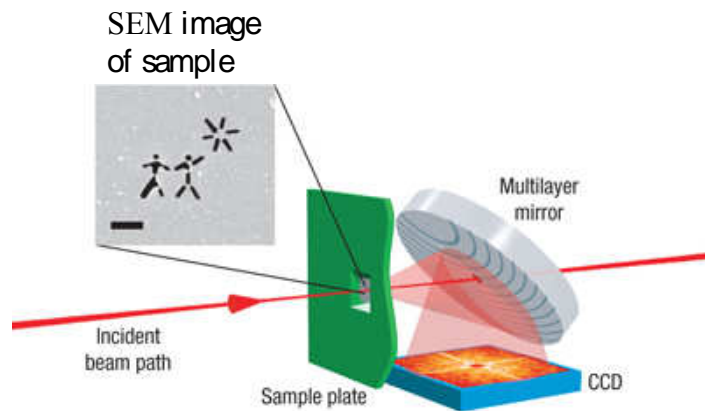


- !! In the year 2000 the “diffraction before destruction” principle was proposed
- !! Effect of ultrashort (2 fs), high-intensity (12 keV) X-ray pulses on T4 Lysozyme was simulated
- !! The simulation was continued for 50 fs after the pulse, the molecule explodes, but the structural information has been gathered.

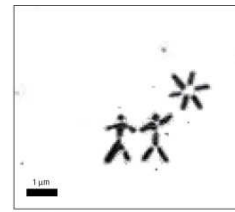
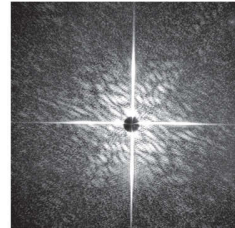


# First experimental demonstration of the “diffraction before destruction” principle

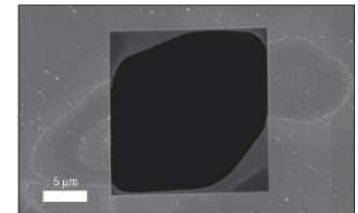
Demonstrated at the first soft-X-ray free electron laser in the world (FLASH facility at DESY), published in 2006



Diffraction pattern

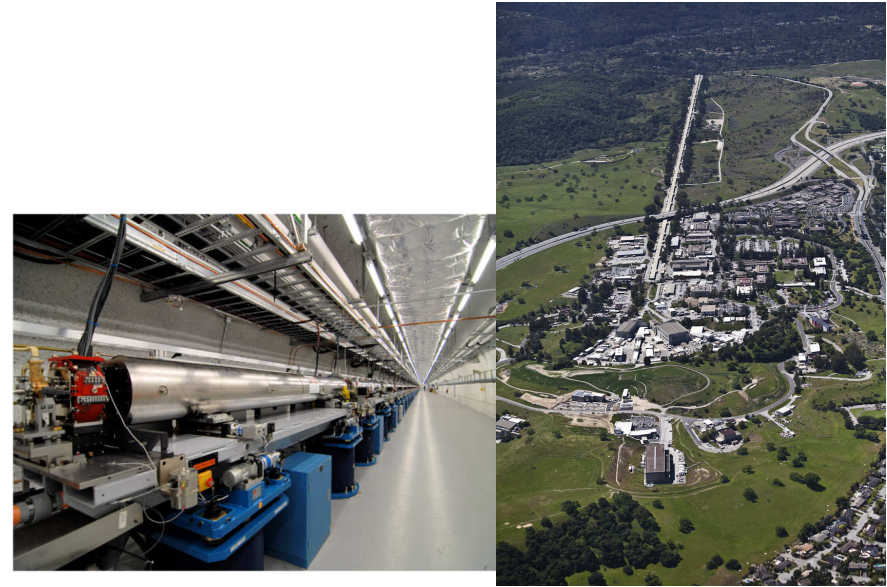
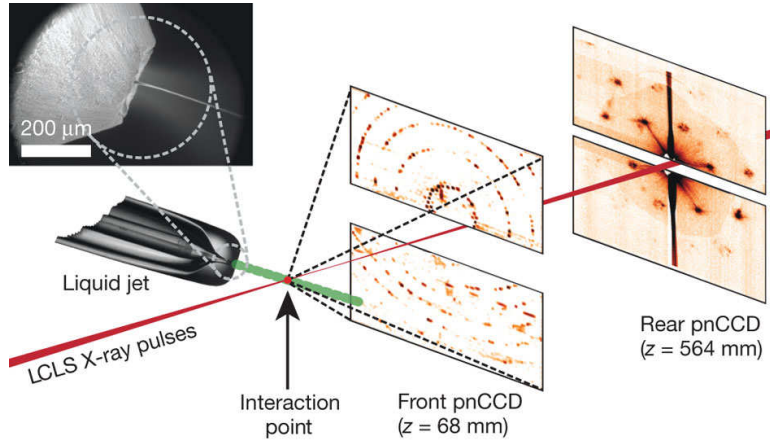


Reconstructed image



SEM image of sample after exposure to the FEL beam

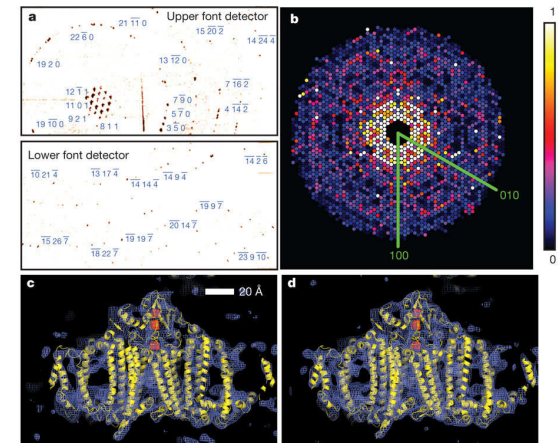
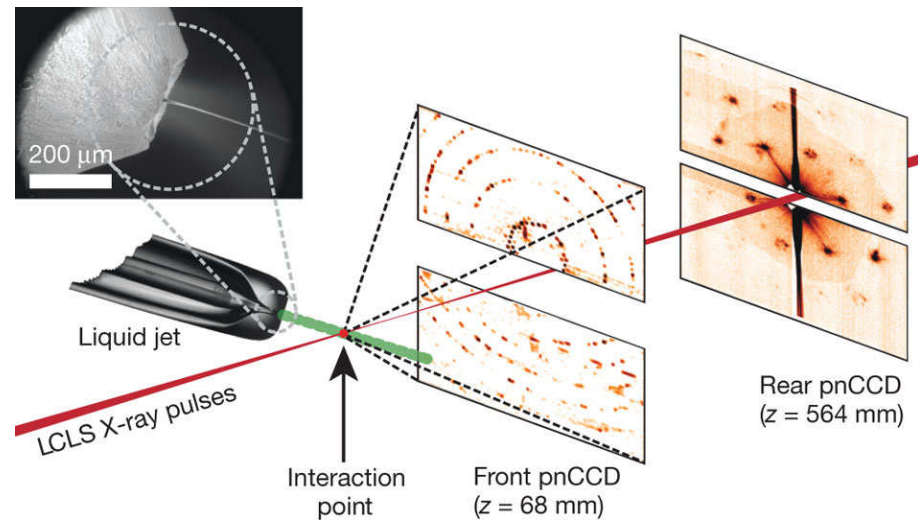
# First demonstration of serial femtosecond X-ray protein nanocrystallography



- !! At the hard-X-ray free-electron laser, the LCLS (Stanford University, DOE)
- !! Data collected from a fully hydrated stream of protein nanocrystals
- !! Two detectors (X-ray p-n junction charge-coupled device (pnCCD))
- !! Structure from Photosystem I (1 MDa membrane protein complex)

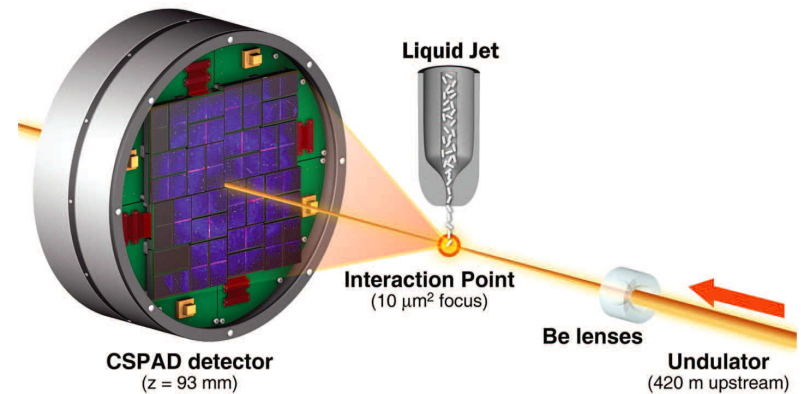
# First demonstration of serial femtosecond X-ray protein nanocrystallography

- !! Photon energy of X-ray pulse 1.8 keV (6.9 Å)
- !!  $10^{12}$  photons per pulse at the sample
- !! pulse durations 10, 70, and 200 fs
- !! Data recorded with 30 Hz (1800 patterns per minute)
- !! Liquid jet with nanocrystals (0.2 to 2 μm)
- !! Crystal hit rate 20%
- !! 1 850 000 X-ray pulses needed
- !! Diffraction patterns with 'partial' Bragg reflections analyzed with Monte Carlo approach



# Serial femtosecond X-ray protein nanocrystallography

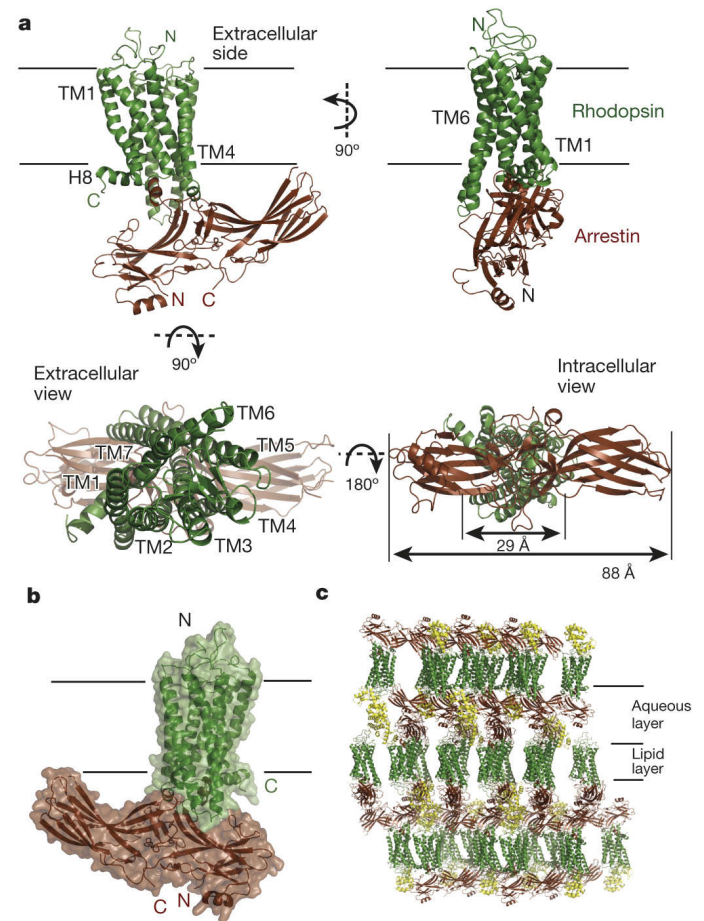
- !! Photon energy of X-ray pulse  
9.5 keV (1.3 Å)
- !!  $10^{11}$  to  $10^{10}$  photons per pulse at the sample
- !! Pulse duration 40 fs or less
- !! Cornell-SLAC pixel array detector (CSPAD)
- !! Data recorded with 120 Hz (7200 patterns per minute)
- !! Resolution down to 1.9 Å





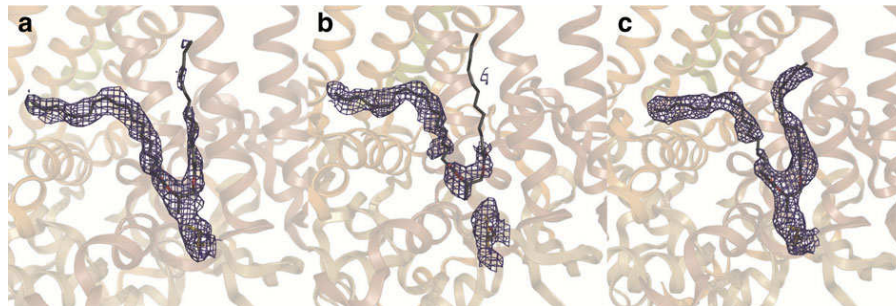
# Crystal structure of rhodopsin bound to arrestin by femtosecond X-ray laser

- !! 12 h of run time
- !! over 5 million detector frames
- !! 22,262 hits were found by the Cheetah hit-finding software
- !! Diffraction patterns from 18,874 crystals indexed and integrated using CrystFEL
- !! Structure with 3.8 Å resolution
- !! Only 6–8 Å at synchrotron sources



# 'Radiation damage free' structures with XFEL

Thioether covalent bond  
in photosynthetic reaction center from *Blastochloris viridis*

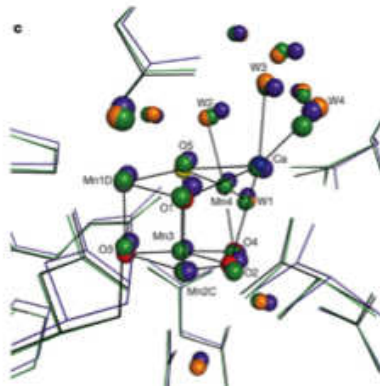


Cryogenic  
temperatures  
(4.4MGy)

Cryogenic  
temperatures  
(77MGy)

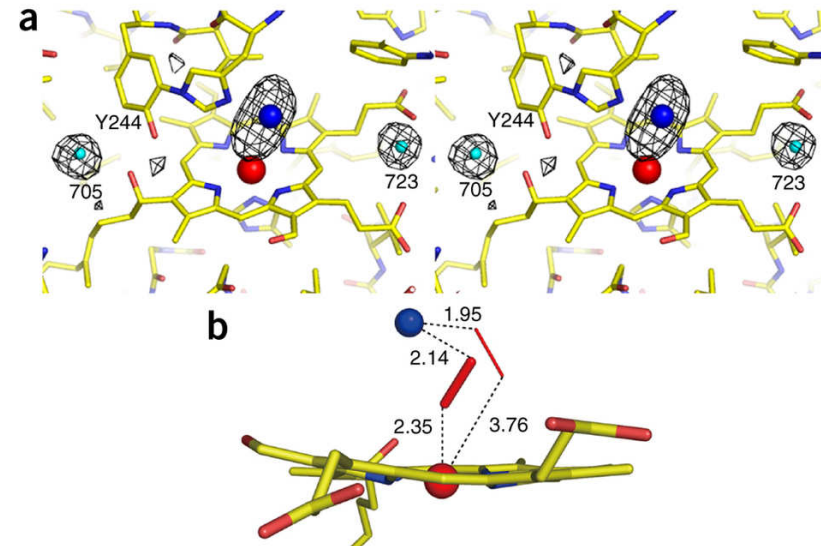
Room temperature  
SFX structure  
(33MGy)

Oxygen evolving complex of  
Photosystem II



Overlay of structures obtained by  
XFEL and synchrotron radiation

The O<sub>2</sub>-reduction site in bovine heart  
cytochrome c oxidase free of radiation damage



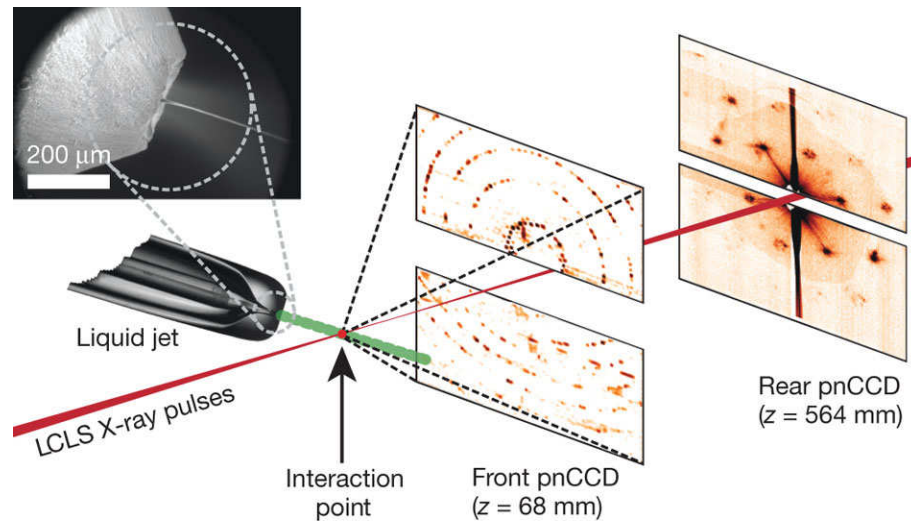
Large single crystal exposed to femtosecond pulses of an X-ray  
free-electron laser (1.9-Å resolution)

Johansson, L. C. et al (2013) Nature Communications 4:2911

Hirata, K. et al (2014) Nature Methods 11, 734–736.

Suga, M. et al (2015) Nature 517, 99-103.

# Serial femtosecond X-ray protein nanocrystallography



- !! Overcomes the radiation damage problem
- !! Room temperature data collection
- !! Fast crystal optimization
- !! Time resolved data collection



# Nanocrystal characterization



# Nanocrystal characterization

Nanocrystals are too small to be detected with an optical microscope



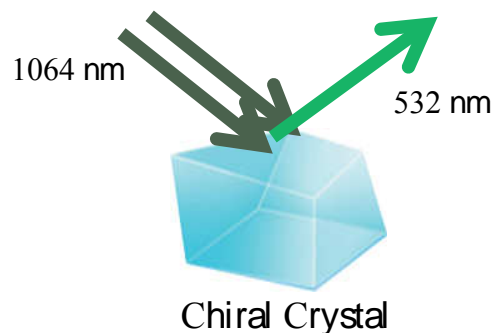
crystal growth cannot be monitored with common methods used in crystallography

Methods of choice:

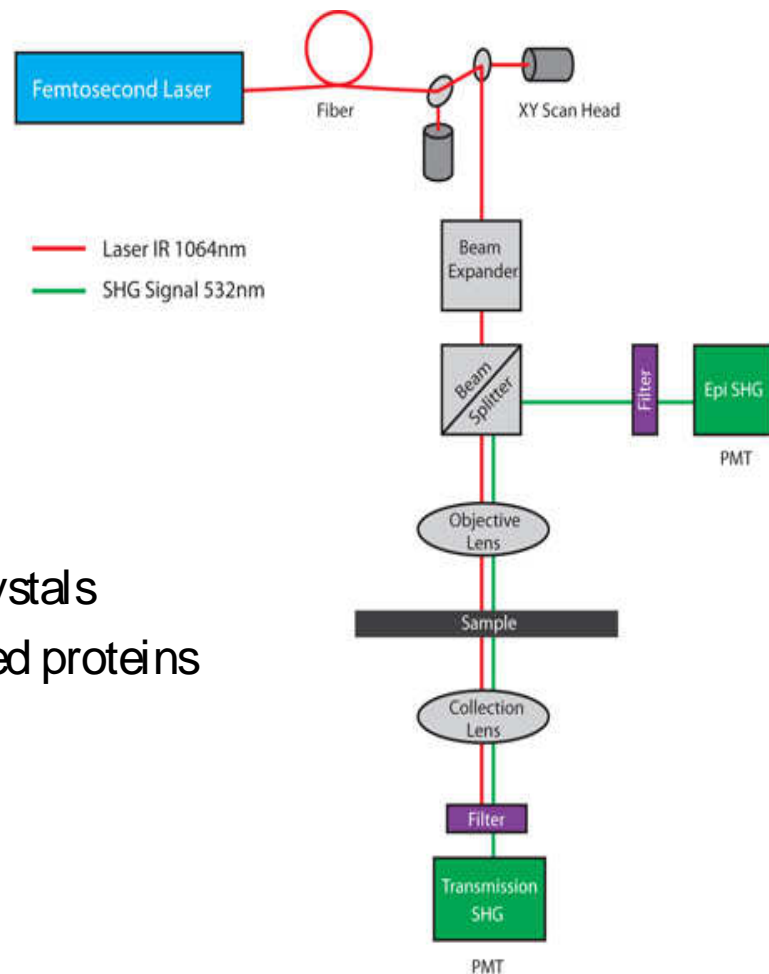
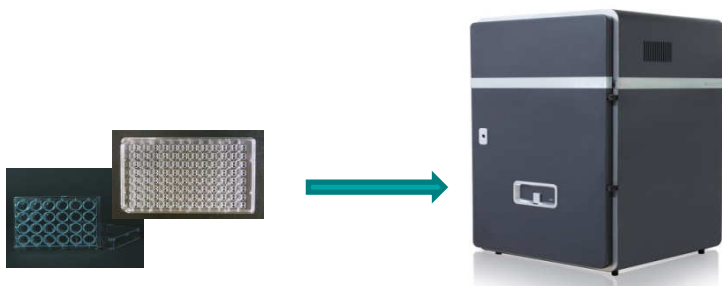
- !! Second order non-linear optical imaging of chiral crystals (SONICC)
- !! Dynamic Light Scattering (DLS)
- !! Nanoparticle Tracking Analysis (NTA)

# SONICC –

## Second order non-linear optical imaging of chiral crystals



- !! Principle: Second Harmonic Generation (SHG)
- !! Detects non-centrosymmetric ordered (chiral) crystals
- !! no signal occurring from solubilized or aggregated proteins



# SONICC – Detection of nanocrystals

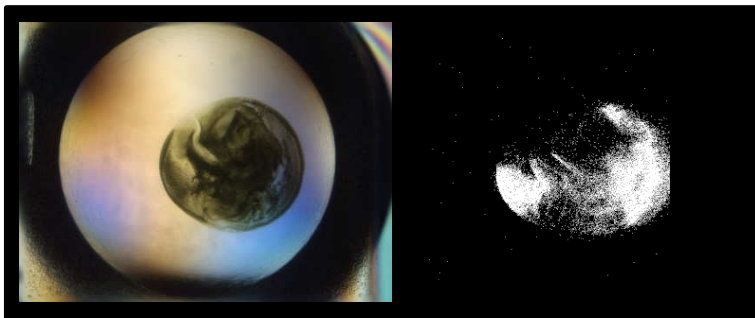
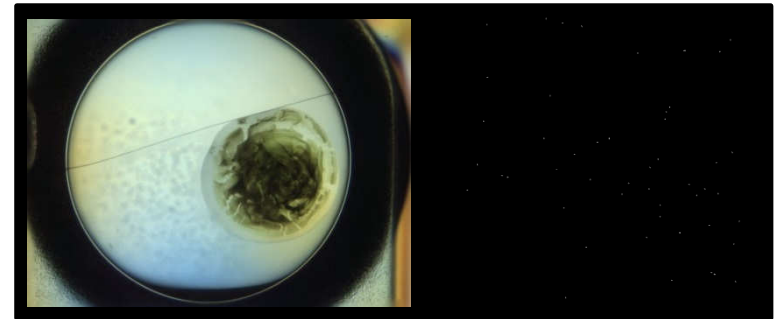
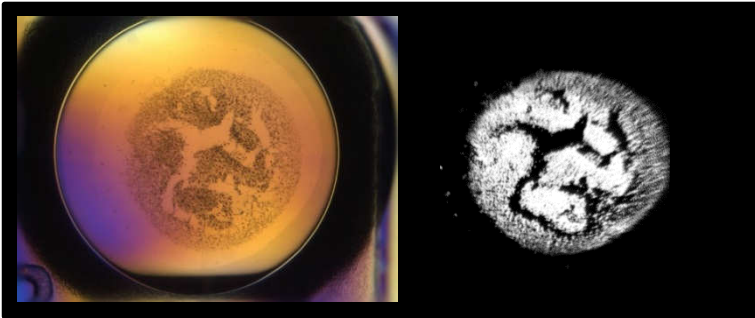
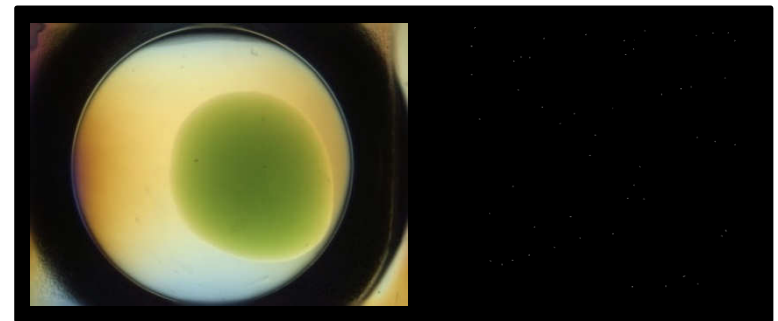
Visible light

SONICC



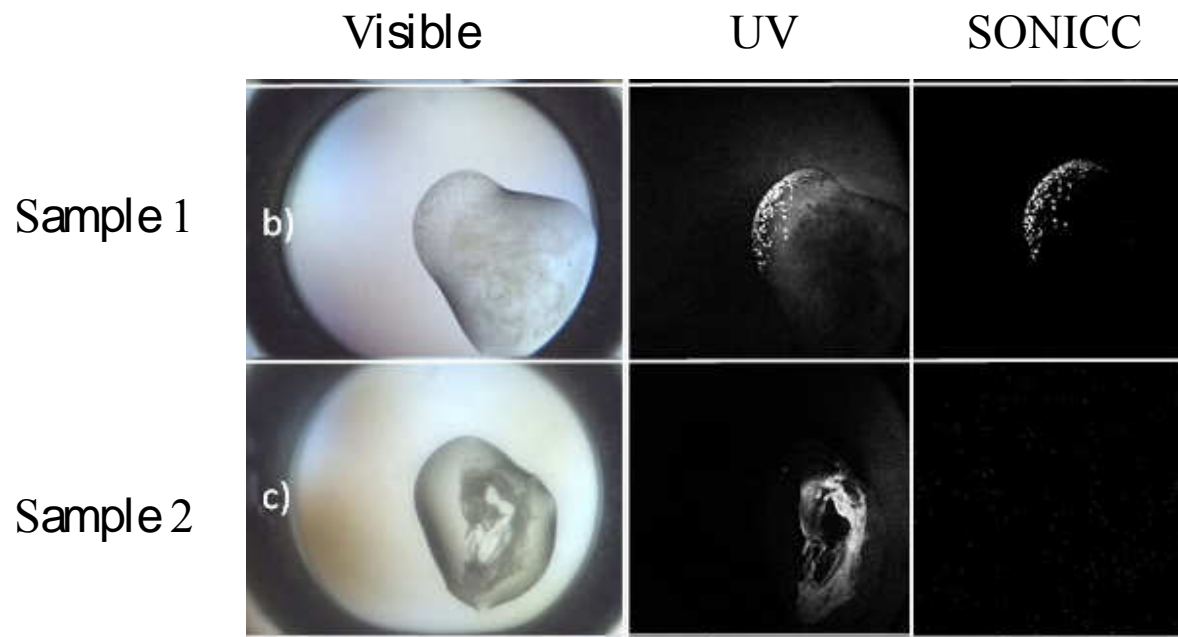
Visible light

SONICC



Powerful technique for  
nanocrystallography

# SONICC – Detection of nanocrystals



# Nonlinear microscopy and principle of Second Harmonic Generation (SHG)

3 phenomena in parallel:

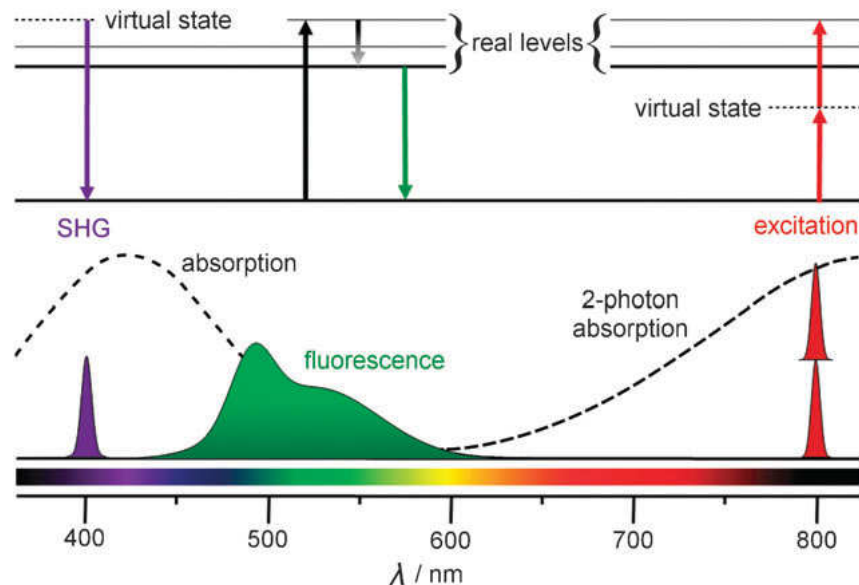
- "! One photon excited fluorescence (1PF)
- "! Two photon excited fluorescence (2PF)
- "! Second harmonic generation (SHG)

1PF and 2PF

- "! Excitation and relaxation in real states

SHG

- "! Parametric process
- "! Scattering through interaction with a virtual states
- "! No excited states, not photo damage
- "! Retains phase, polarization and directional information



# Molecular design for SHG and therefore SONICC

## 1.) Medium without center of inversion

Ensemble of chiral molecules: protein crystals, collagen, dyes in a biological membrane

(No SHG: Random order and centro-symmetric crystals: protein solution, aggregated protein, most of salt crystals)

## 2.) Sufficient intensity of scattered light

"! Intensity depends on crystal size, crystal class, amino acid sequence...

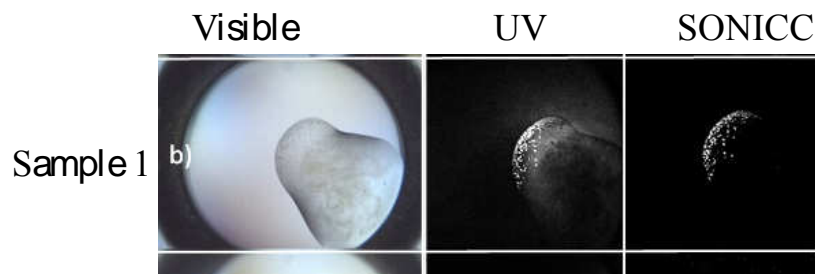
"! "-conjugated systems lead to signal enhancement (resonance enhancement)



Proteins with cofactors like chlorophyll and heme groups, GFP-like proteins, proteins with high density of aromatic amino acids lead to strong SONICC signal

# SONICC

## High symmetry crystal classes not necessarily detectable

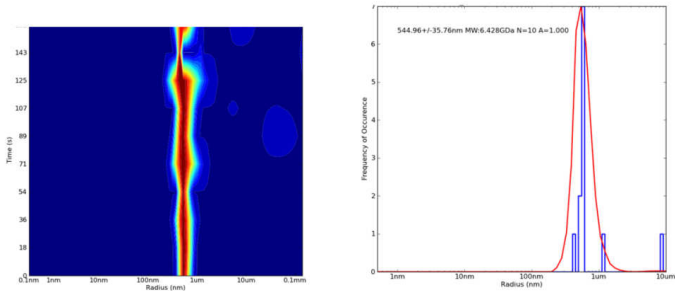
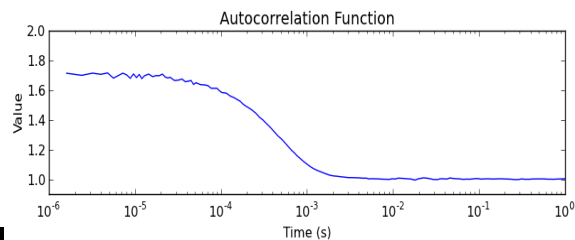
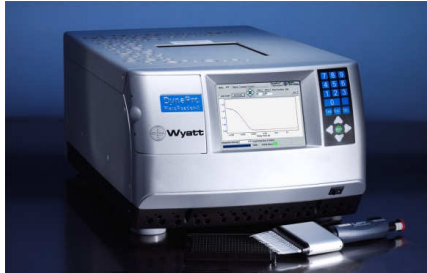


Sample 2

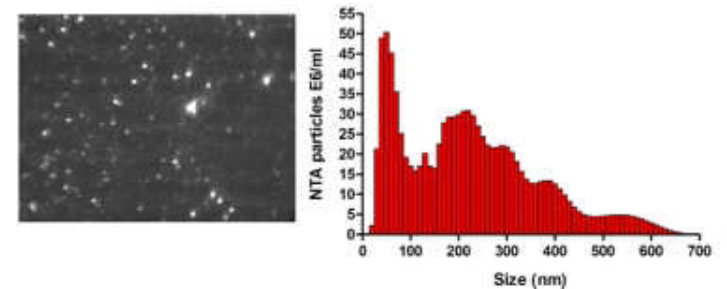
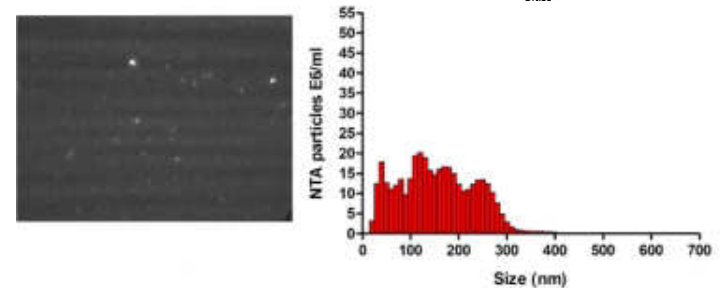
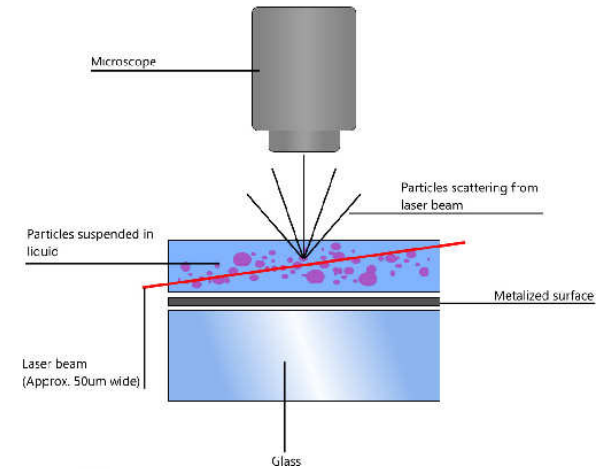
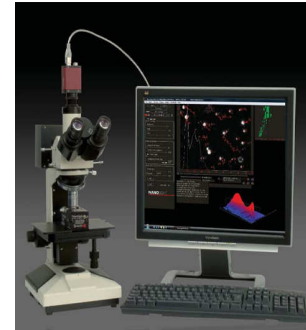
Crystal system	Point group/ Crystal Class		Space groups (international short symbol)	Generates SHG	% of known structures (A)	% of structures in the space group that do not have any tryptophan residues (B)	% of structures with no tryptophan residues (organized by space group) (=A*B)
	Intl	Schönflies					
Triclinic	1	C <sub>1</sub>	P1	yes	4.04	18.34%	0.74%
Monoclinic	2	C <sub>2</sub>	P2, P2 <sub>1</sub> , C2	yes	24.78	16.88%	4.18%
Orthorhombic	222	D <sub>2</sub>	P222, P222 <sub>1</sub> , P2 <sub>1</sub> 2 <sub>1</sub> 2, P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub> , C222 <sub>1</sub> , C222, F222, I222, I2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	yes	36.36	16.83%	6.12%
Trigonal	3	C <sub>3</sub>	P3, P3 <sub>1</sub> , P3 <sub>2</sub> , R3	yes	4.04	20.24%	0.82%
	32	D <sub>3</sub>	P312, P321, P3 <sub>1</sub> 12, P3 <sub>1</sub> 21, P3 <sub>2</sub> 12, P3 <sub>2</sub> 21, R32	yes	7.3	18.18%	1.32%
Tetragonal	4	C <sub>4</sub>	P4, P4 <sub>1</sub> , P4 <sub>2</sub> , P4 <sub>3</sub> , I4, I4 <sub>1</sub>	questionable	2.36	19.26%	0.45%
	422	D <sub>4</sub>	P422, P4 <sub>2</sub> 2, P4 <sub>1</sub> 22, P4 <sub>1</sub> 2 <sub>1</sub> 2, P4 <sub>2</sub> 22, P4 <sub>2</sub> 2 <sub>1</sub> 2, P4 <sub>3</sub> 22, P4 <sub>3</sub> 2 <sub>1</sub> 2, I422, I4 <sub>1</sub> 22	questionable	10.02	17.33%	1.73%
Hexagonal	6	C <sub>6</sub>	P6, P6 <sub>1</sub> , P6 <sub>5</sub> , P6 <sub>2</sub> , P6 <sub>4</sub> , P6 <sub>3</sub>	questionable	3.6	19.46%	0.70%
	622	D <sub>6</sub>	P622, P6 <sub>2</sub> 22, P6 <sub>2</sub> 22, P6 <sub>2</sub> 22, P6 <sub>4</sub> 22, P6 <sub>5</sub> 22	questionable	4.26	20.07%	0.85%
Cubic	23	T	P23, F23, I23, P2 <sub>1</sub> 3, I2 <sub>1</sub> 3	questionable	1.22	19.88%	0.24%
	432	O	P432, P4 <sub>3</sub> 2, F432, F4 <sub>1</sub> 32, I432, P4 <sub>3</sub> 32, P4 <sub>1</sub> 32, I4 <sub>1</sub> 32	questionable	.59	25.85%	0.15%

# Size distribution and homogeneity of nanocrystals

## Dynamic light scattering (DLS)

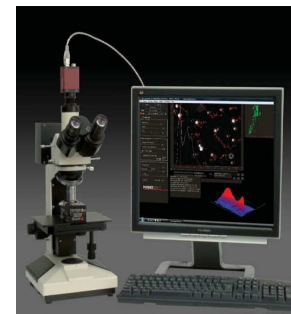


## Nanoparticle Tracking Analysis (NTA)





# Size distribution and homogeneity of nanocrystals



	DLS	NTA by NanoSight
Size range	About 1 – 1000 nm	About 30 – 1000 nm
Size accuracy	Accurate for monodisperse samples, inaccurate for polydisperse samples	Accurate for monodisperse and polydisperse samples
Peak resolution	Low (>3 fold difference in diameter)	High (<0.5 fold difference in diameter)
Contaminations	Large particles problematic	Large particles have little influence
Reproducibility	more	less
Duration of measurement	2 – 5 min	5 min to 1 hour

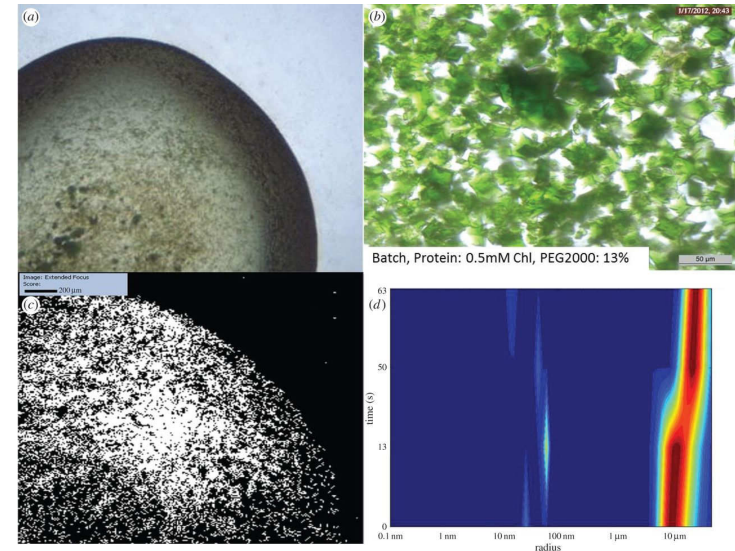
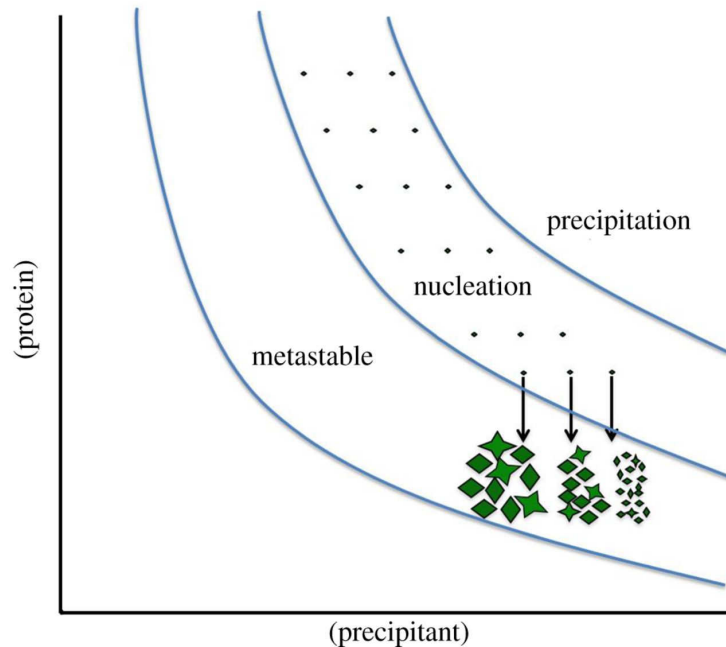


# Nanocrystal growth

# Nanocrystal growth

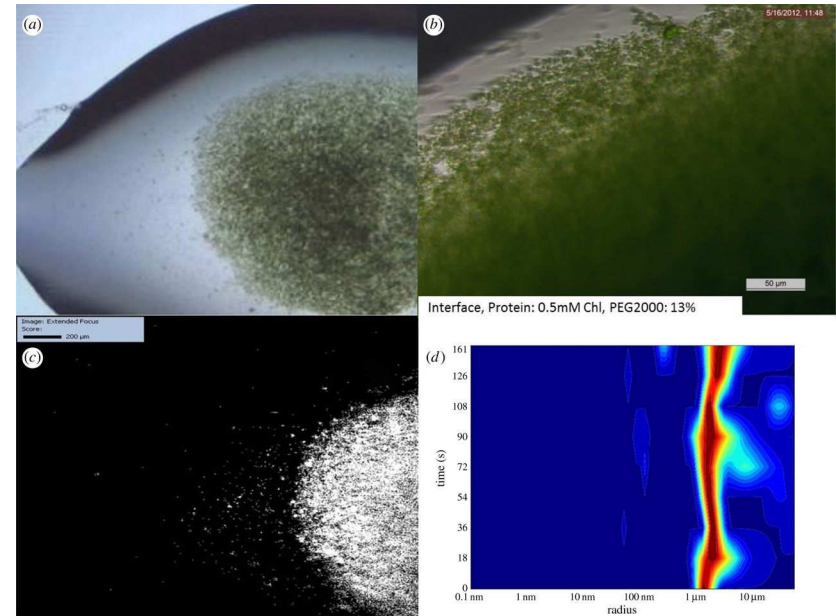
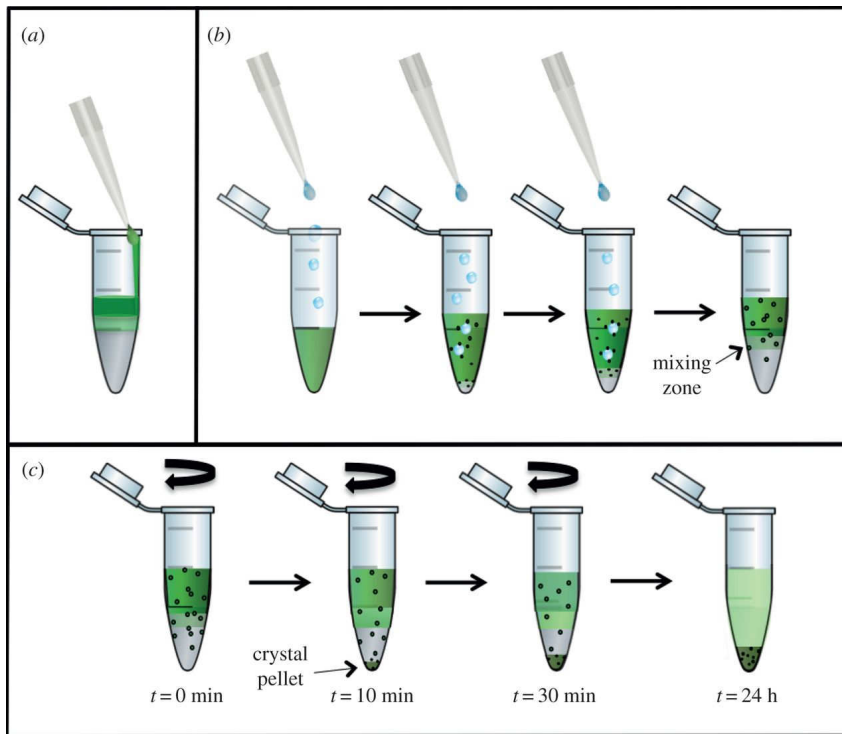
- !! Batch method
- !! Free interface diffusion (FID)
- !! Free interface diffusion with centrifugation

# Nanocrystal growth - Batch method



- !! Phase diagram should be known
- !! Rapid mixing of protein and precipitation buffer (Eppendorf tube, Falcon tube)
- !! Final concentration should be in nucleation zone
- !! May lead to polycrystalline samples

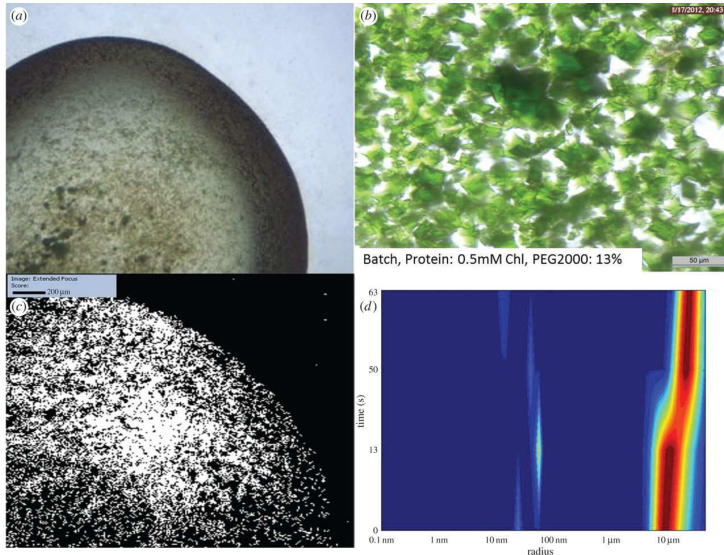
# Nanocrystal growth - Free interface diffusion (FID)



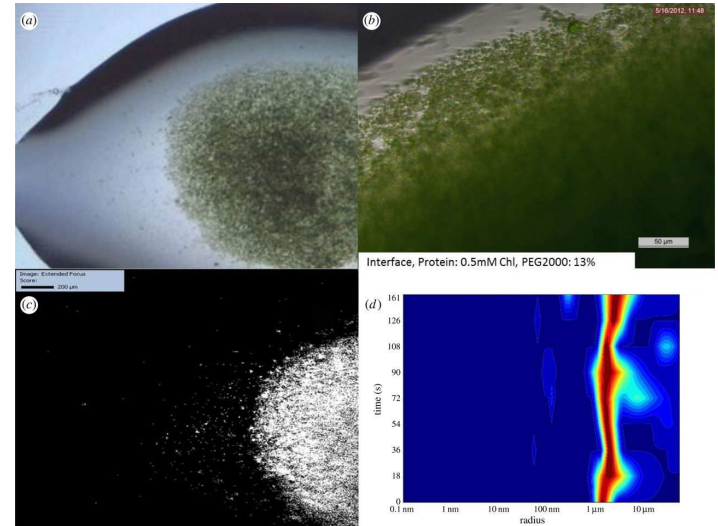
- !! Crystal grow at interface of high concentrated protein solution and precipitant buffer
- !! Precipitant solution drop wise added to protein solution
- !! Process can be accelerated by centrifugation

# Nanocrystal growth

## Batch method

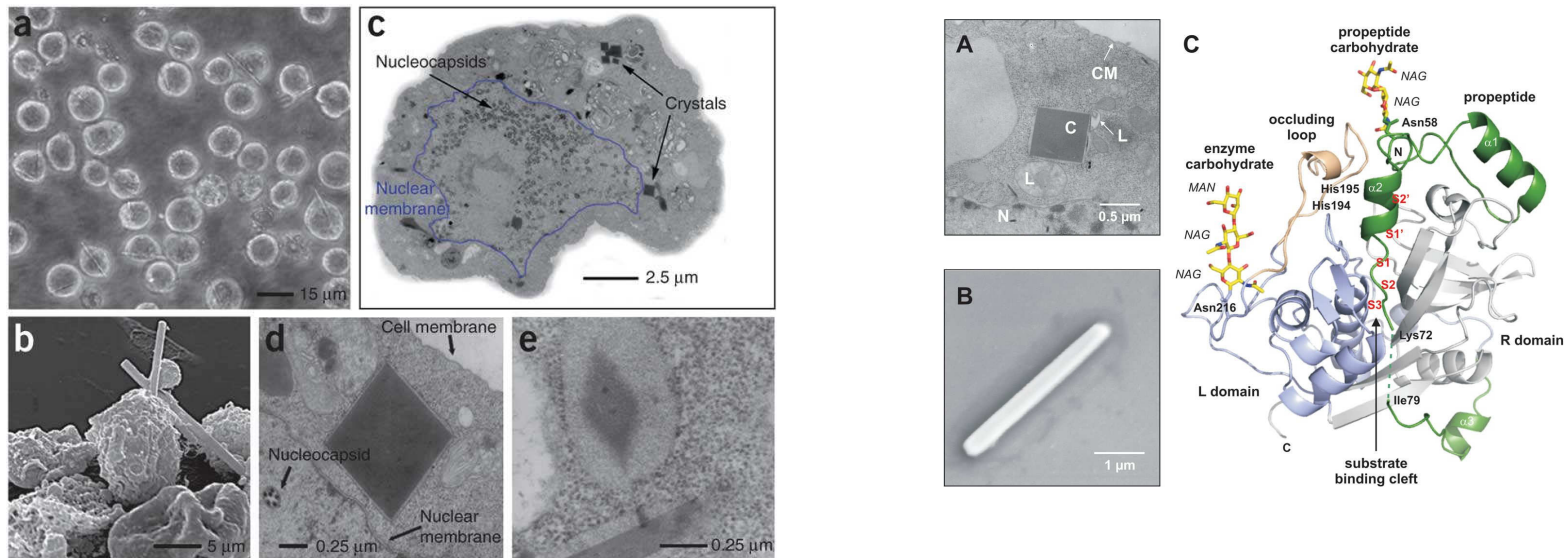


## FID method



- !! Quenching of crystal growth may be necessary
- !! Transport of nanocrystals can be problematic
- !! Crystallization on site (LCLS, SACLA)

# In vivo crystallization to generate nanocrystals



- !! Cysteine protease cathepsin from *Trypanosoma brucei* (TbCatB) crystals within Sf9 insect cells
- !! TbCatB is promising target for treatment of sleeping sickness
- !! Crystals too small for synchrotron-based experiments
- !! With SFX structure was solved with 2.1 Å resolution
- !! In the future: Imaging of crystals in cells?



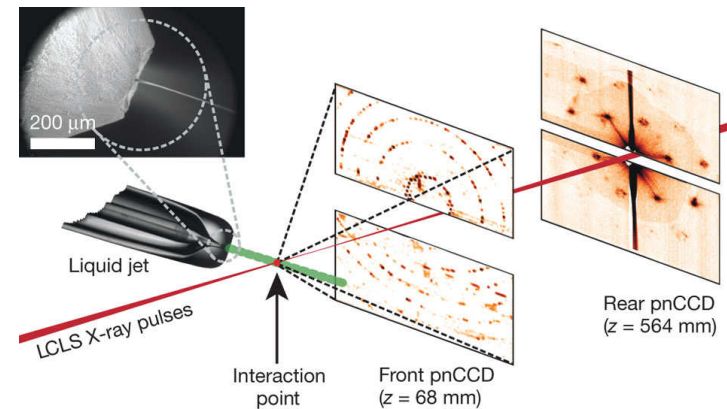
# Nanocrystal injection/delivery methods



# Nanocrystal injection methods

## Requirements

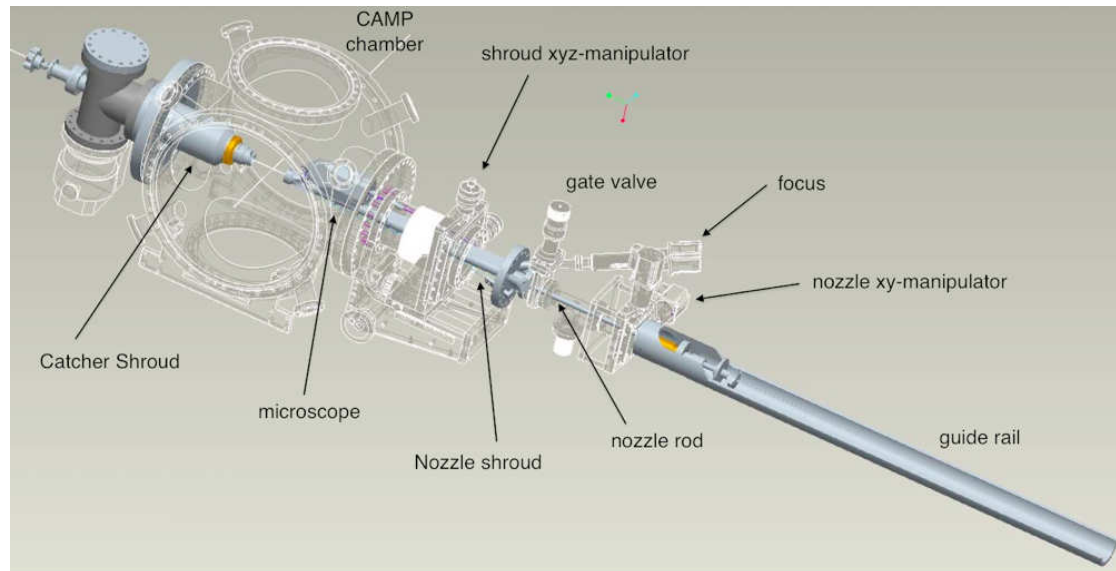
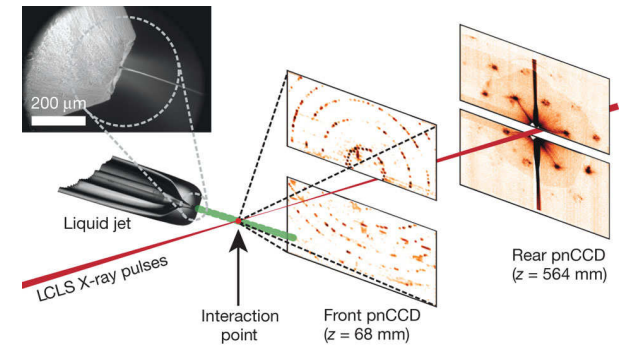
- !! Stable running jet
- !! No clogging of system
- !! No dissolving of crystals
- !! Low sample consumption
- !! Low background scattering



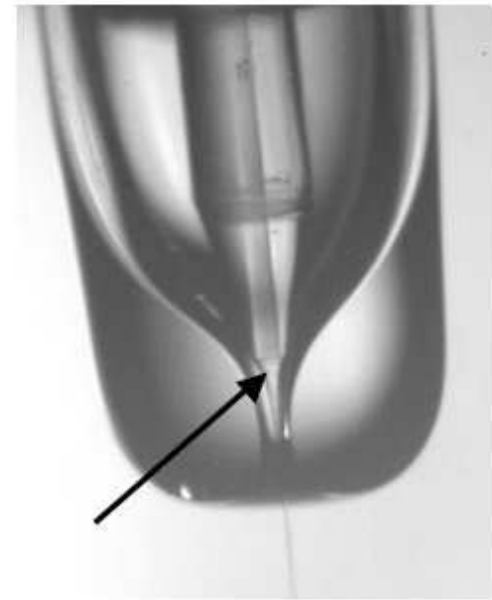
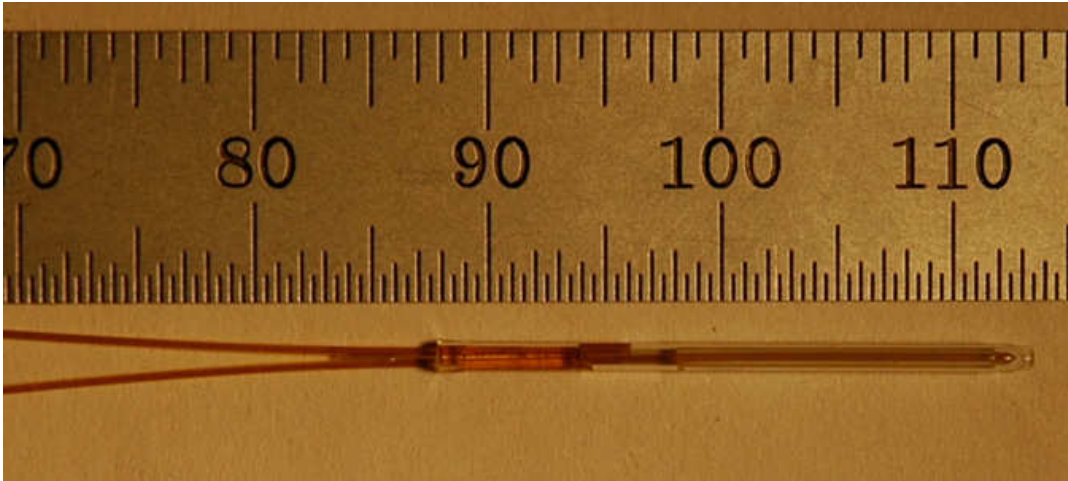
Chapman, H. N. et al (2011) Nature 470, 73-77.

# Injection of crystal into X-ray beam

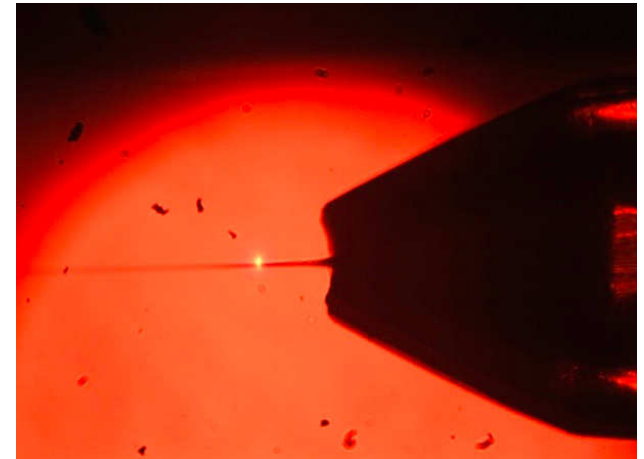
- !! Liquid jet injector
- !! Nozzle shroud and catcher shroud
- !! Based on Gas Dynamic Virtual Nozzle (GDVN)



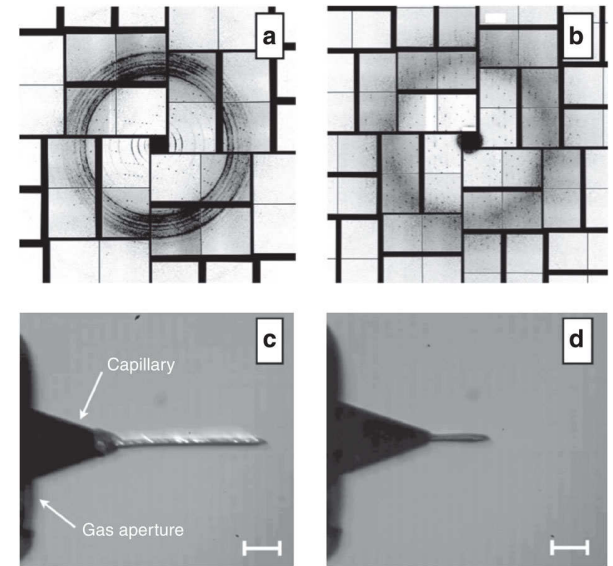
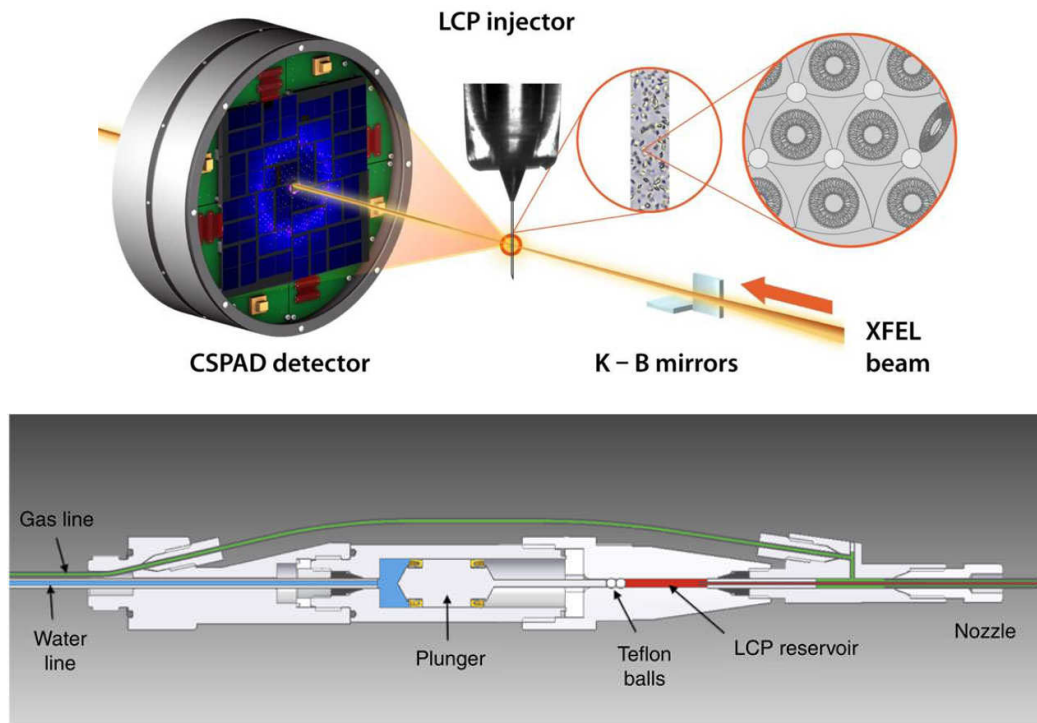
# Gas Dynamic Virtual Nozzle (GDVN)



- !! Inner capillary (20 – 50  $\mu\text{m}$ ) with liquid crystal stream
- !! Co-flowing gas in an outer capillary reduce the diameter of the liquid jet
- !! Resulting jet diameter down to 4  $\mu\text{m}$
- !! Flow rates 10 to 20  $\mu\text{L min}^{-1}$



# Lipid Cubic Phase (LCP) injector

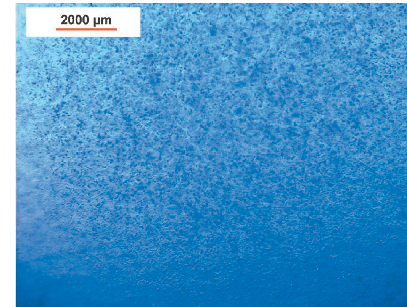
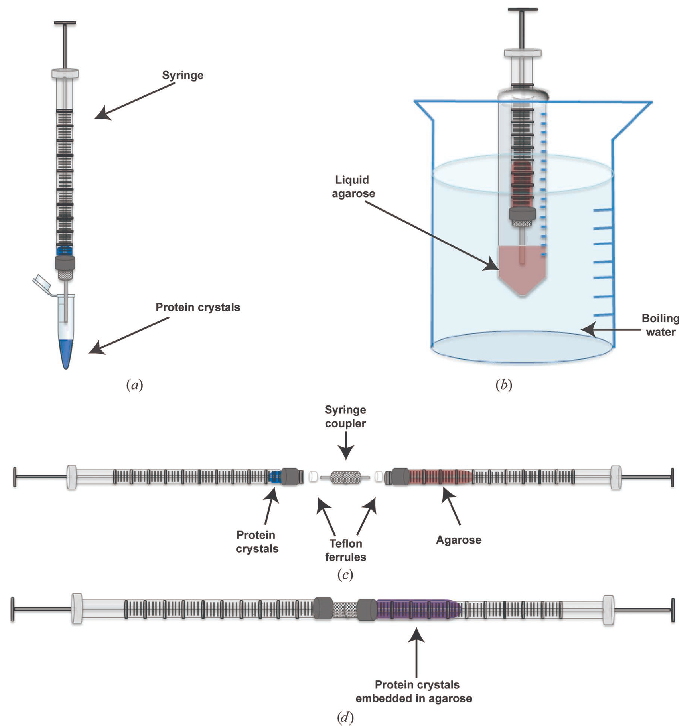


- !! Injection of gel-like LCP with microcrystals
- !! 10–50  $\mu\text{m}$  diameter stream
- !! Protein consumption reduced by a factor of 20 compared with to GDVN nozzle

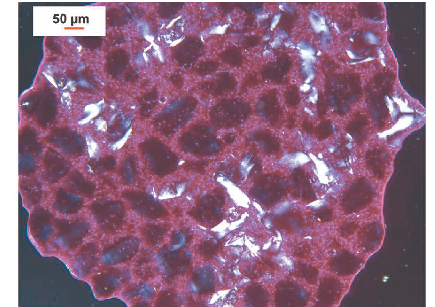
Weierstall, U. et al. (2014) Nat. Commun. 5, 3309.

Liu, W. et al (2014) Science 342, 1521-1524.

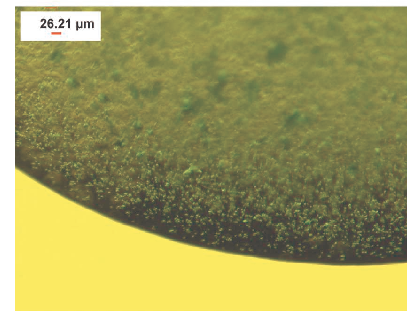
# Agarose as injection medium



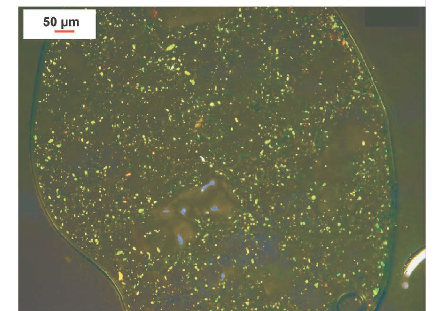
(a)



(b)



(c)

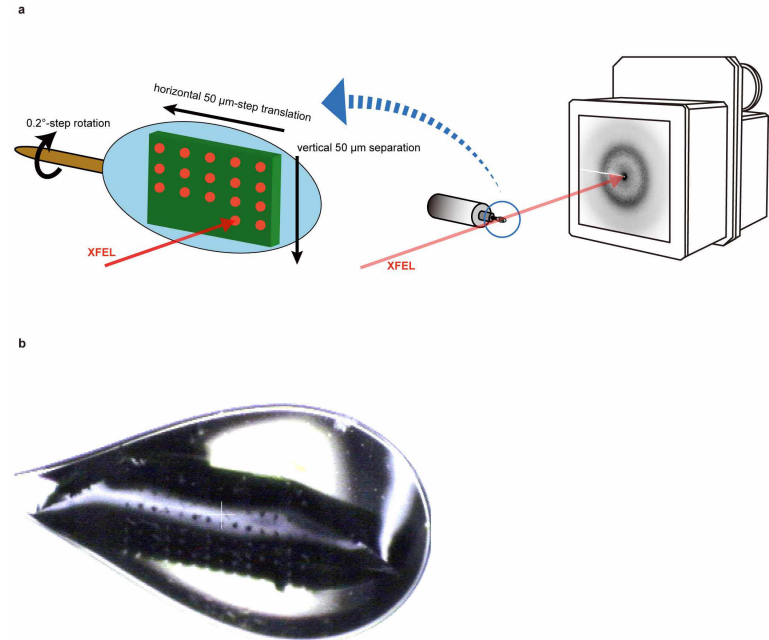


(d)

- !! Injection with LCP injector
- !! Microcrystallization via conventional methods and then mixed with agarose
- !! Protein consumption reduced by a factor of 100 compared with the GDVN nozzle

# Fixed-target delivery methods – Rotating large crystal

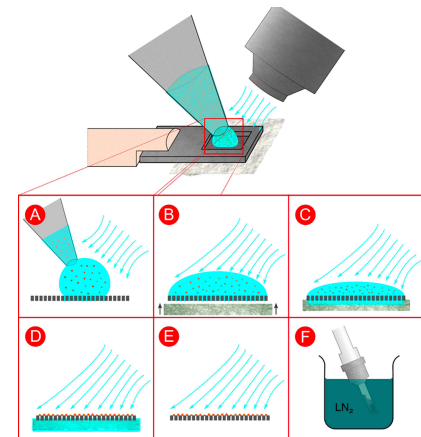
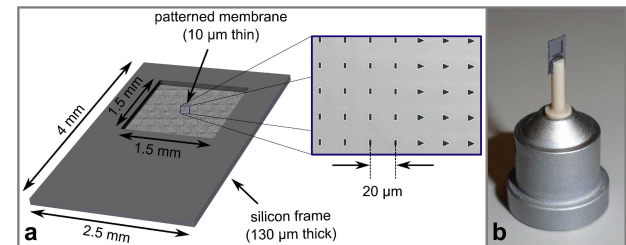
- !! Developed at SACLA in Japan
- !! Large single crystal ( $\mu\text{m}/\text{mm}$  range) rotated stepwise in order to record still diffraction images
- !! Hundreds of crystals “scanned” with the X-ray laser
- !! High resolution, radiation damage-free structures of Photosystem II and Cytochrome C oxidase





# Fixed-target delivery methods – Silicon chips

- !! Microcrystal slurry is applied on silicon chip with micropores
- !! Chip is scanned with x-ray beam
- !! Room temperature and cryogenic temperatures
- !! Quick and easy sample preparation

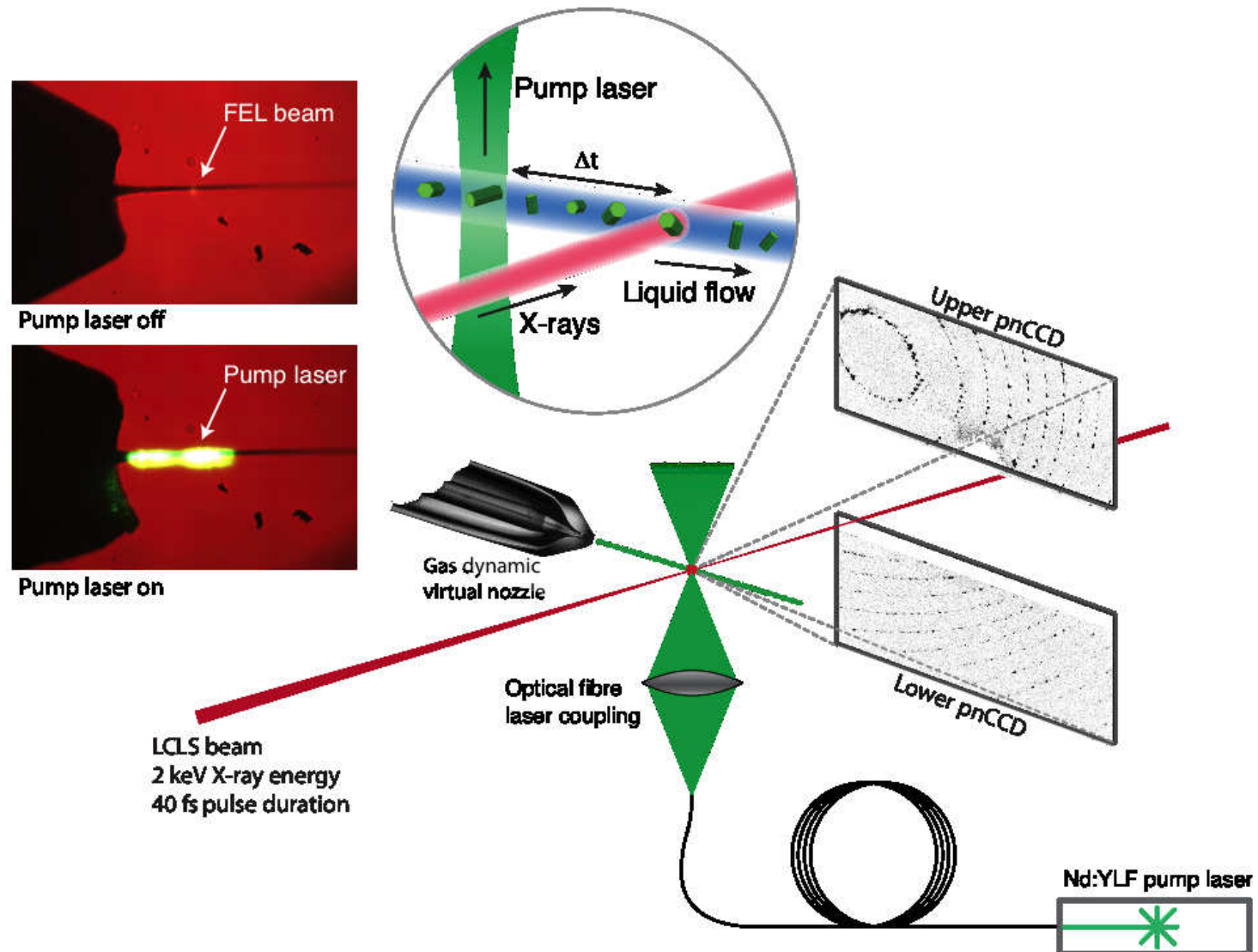


# Time-resolved studies

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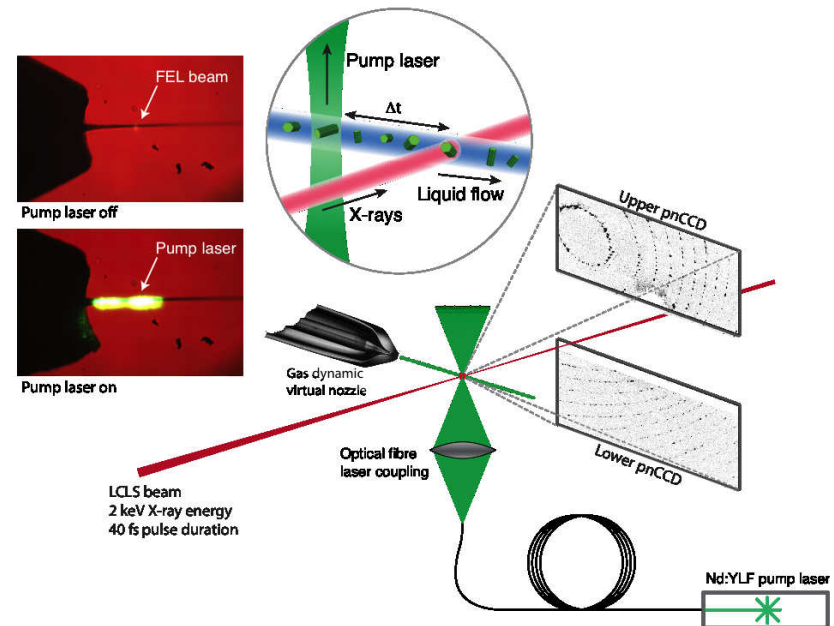


# First time-resolved studies at XFEL on Photosystem I

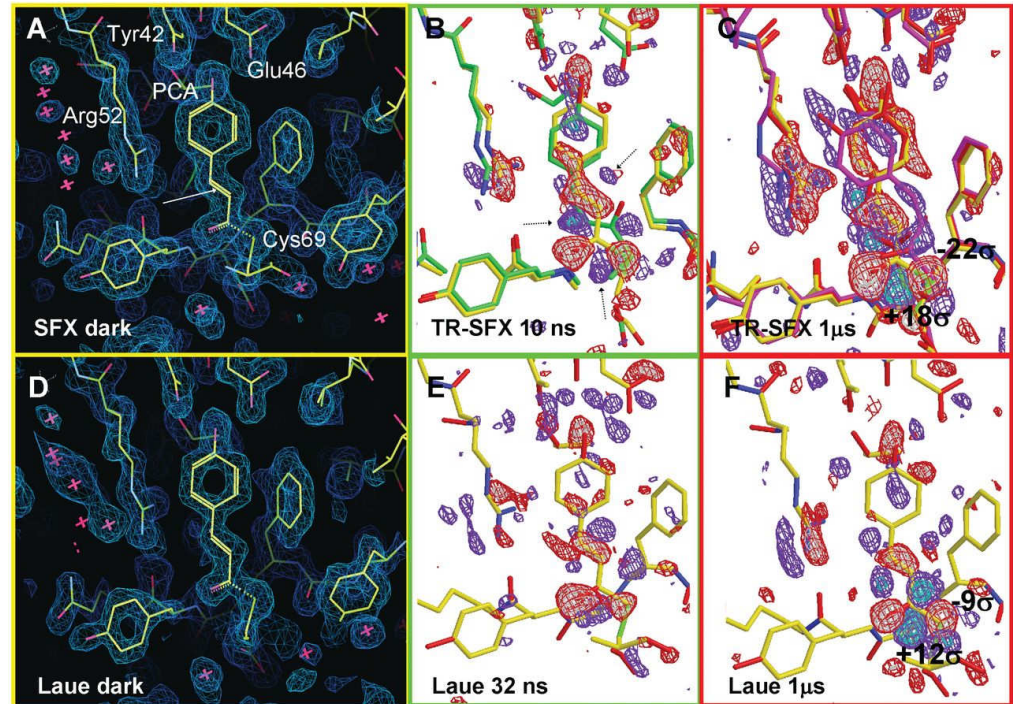
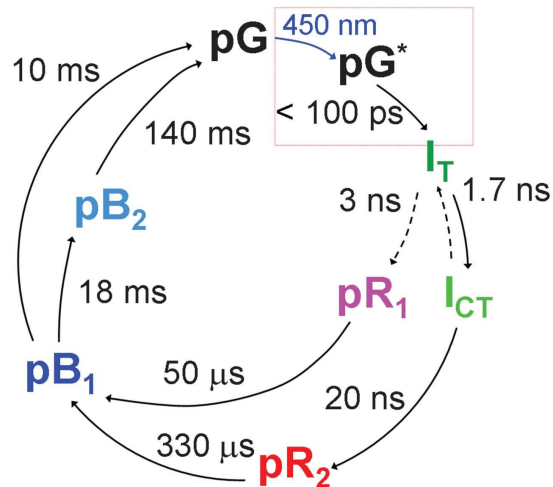
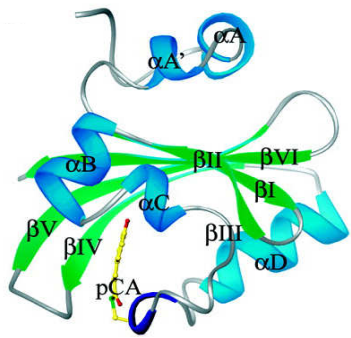


# First time-resolved studies at XFEL on Photosystem I

- !! XFEL synchronized with an optical pump laser
- !! Light-induced changes of Photosystem I-Ferredoxin co-crystals observed at time delays of 5 to 10  $\mu\text{s}$  after excitation
- !! Enables study of the dynamics in hydrated single nano- to micron-sized crystals

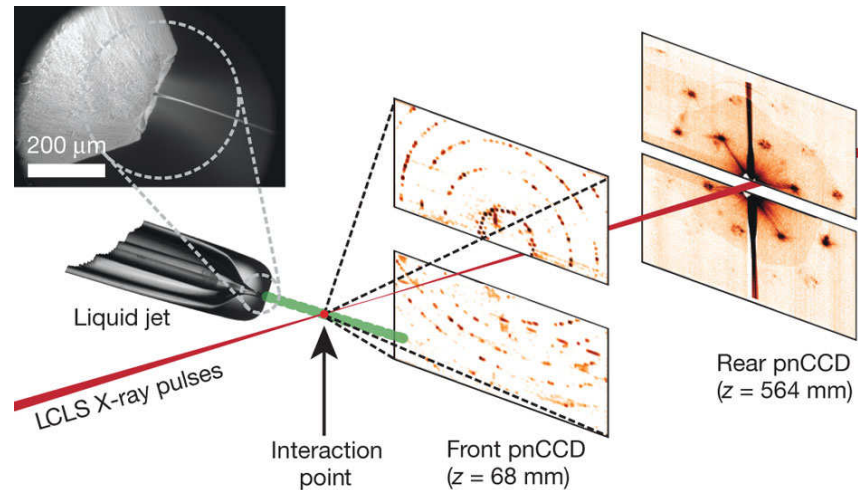


# Time-resolved studies on photoactive yellow protein (PYP)



- !! Light-triggered dynamics in PYP (pump probe experiments)
- !! High-resolution (1.6 Å), time-resolved difference electron density maps
- !! Study of biological reactions on fs time scale possible

# Serial femtosecond X-ray protein nanocrystallography - Outlook



Chapman, H. N. et al (2011) Nature 470, 73-77.

- !! Phasing problem
- !! Carefully study radiation damage
- !! Time consumption
- !! Accessibility
- !! Mixing of proteins with substrates
- !! ...



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Chapman group

Fromme group

Kärtner group

CFEL team

ASU team



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THANK YOU!

